Na\(^+\) Interaction with the Pore of Shaker B K\(^+\) Channels: Zero and Low K\(^+\) Conditions

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**ABSTRACT** The Shaker B K\(^+\) conductance (G\(_K\)) collapses (in a reversible manner) if the membrane is depolarized and then repolarized in, 0 K\(^+\), Na\(^+\)-containing solutions (Gómez-Lagunas, F. 1997. *J. Physiol.* 499:3-15; Gómez-Lagunas, F. 1999. *Biophys. J.* 77:2988-2998). In this work, the role of Na\(^+\) ions in the collapse of G\(_K\) in 0-K\(^+\) solutions, and in the behavior of the channels in low K\(^+\), was studied. The main findings are as follows. First, in 0-K\(^+\) solutions, the presence of Na\(^+\) ions is an important factor that speeds the collapse of G\(_K\). Second, external Na\(^+\) fosters the drop of G\(_K\) by binding to a site with a K\(_d\) = 3.3 mM. External K\(^+\) competes, in a mutually exclusive manner, with Na\(^+\) for binding to this site, with an estimated K\(_d\) = 80 \(\mu\)M. Third, NMG and choline are relatively inert regarding the stability of G\(_K\); fourth, with [K\(^+\)] = 0, the energy required to relieve Na\(^+\) block of Shaker (French, R.J., and J.B. Wells. 1977. *J. Gen. Physiol.* 70:707–724; Starkus, J.G., L. Kuschel, M. Rayner, and S. Heinemann. 2000. *J. Gen. Physiol.* 110:539–550) decreases with the molar fraction of Na\(^+\) (X\(_{Na}\)), in an extent not accounted for by the change in \(\Delta\mu_{Na}\). Finally, when X\(_{Na}\) = 1, G\(_K\) collapses by the binding of Na\(^+\) to two sites, with apparent K\(_s\) of 2 and 14.3 mM.

**KEY WORDS:** K\(^+\) affinity • Na\(^+\) block • conductance • selectivity • zero K\(^+\)

**INTRODUCTION**

In addition to permeate through voltage-dependent K channels (Kv channels)* and modulate their gating, K\(^+\) ions are an essential factor needed to keep these proteins in their normal, functional, state. It is now becoming clear however, that the K\(^+\) requirements of Kv channels are highly variable, as revealed by the diversity of effects that K\(^+\) depletion exerts on different kinds of channels. For example, in the absence of K\(^+\) on both sides of the membrane, the activity of the delayed rectifier (DR) squid K channel is irreversibly lost (Chandler and Meves, 1970; Almers and Armstrong, 1980; Khodorov et al., 1997). On the other hand, both DR channels of bullfrog sympathetic neurons (Block and Jones, 1997) as well as mammalian DRs, of the Kv2.1 subfamily, remain active in 0 K\(^+\), allowing a substantial permeation of Na\(^+\), but becoming anomalously immune to the addition of TEA (Zhu, and Ikeda, 1993; Ikeda and Korn, 1995). Similarly, Kv1.5 channels also remain active in 0 K\(^+\), although they only allow a small, fast inactivating, and TEA-sensitive, flux of Na\(^+\) through them (Zhurenko et al., 2000). In contrast, other Kv channels are apparently insensitive to K\(^+\) depletion. For example, Kv1.3 channels, do not allow a measurable permeation of Na\(^+\), and remain active after the exposure to 0 K\(^+\) (Immke et al., 1998; see also Jäger et al., 1998).

Recently, it was shown that in the absence of K\(^+\) on both sides of the membrane, the Shaker B K\(^+\) conductance (G\(_K\)) collapses when the membrane is depolarized and then repolarized. Briefly, Gómez-Lagunas (1997, 1999) reported the following. First, G\(_K\) collapses if the channels are gated, by the delivery of standard activating pulses, while they are immersed in Na\(^+\)-containing, 0 K\(^+\), solutions. The extent of collapse depends on the number of pulses but not on their frequency. Second, in contrast, the halt in the conductance is completely prevented if the channels are kept closed (are not gated) while they are in 0 K\(^+\). Third, depolarized holding potentials (HP, above −50 mV) impede the drop of G\(_K\). Fourth, external K\(^+\) protects G\(_K\) by binding to a site in the channels with rather low, millimolar, affinity (apparent K\(_d\) = 2.9 mM). Fifth, among divalent cations only Ba\(^{2+}\) is able to replace K\(^+\), protecting G\(_K\); and finally, very prolonged depolarizations (seconds to minutes) recover the, previously lost, K\(^+\) conductance.

The above observations were interpreted as meaning that closing without K\(^+\) sinks the channels into a stable nonconducting, nonactivated, closed state(s) (Gómez-Lagunas, 1997). It seems that the channels need to close with a K\(^+\) ion(s) bound to them, otherwise they sink into a reluctant conformation where they remain unable to

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*Abbreviations used in this paper: DR, delayed rectifier; HP, holding potential; Kv channel, voltage-dependent K channel; m.c, maximal collapse.
conduct K\(^+\), no matter how long Vm is kept at the HP or hyperpolarized potentials (Gómez-Lagunas, 1997, 1999; Melishchuk et al., 1998). Looking at the selectivity of the site(s) involved in the drop of G\(_K\), preliminary observations have indicated that the presence of Na\(^+\) ions might significantly foster the collapse of G\(_K\) in 0-K\(^+\) solutions, probably by promoting the displacement of K\(^+\) from the pertinent site(s) in the channels (Gómez-Lagunas, 1997; Melishchuk et al., 1998). Thus, to further understand the mechanism underlying the fall of G\(_K\) in this work the role of Na\(^+\) ions in both the collapse of the Shaker K\(^+\) conductance in 0-K\(^+\) solutions, and in the behavior of the channels in low K\(^+\), is investigated.

Here, it is shown that Na\(^+\) ions foster the collapse of G\(_K\) in the absence of K\(^+\). External K\(^+\) and Na\(^+\) compete, in a mutually exclusive manner, for binding to an externally located site (probably external to the selectivity filter), where G\(_K\) is modulated (available versus reluctant). On the other hand, to see if the collapse of G\(_K\) occurs as a discontinuous change in the properties of the channels in the limit of zero K\(^+\), experiments with internal solutions containing both K\(^+\) and Na\(^+\) ions were done. The results show that as the molar fraction of Na\(^+\) (X\(_{Na}\)) increases, the energy required to relieve Na\(^+\) block changes in an extent not accounted for by the change in the driving force of Na\(^+\). This suggests that, as X\(_{Na}\) increases, there is either an increased electrostatic repulsion between the blocking Na\(^+\) and a neighbor ion, or, more likely, a deformation (change of the energy profile) of the pore, that could be maximal in the limit X\(_{Na}\) = 1. Under the latter conditions, G\(_K\) collapses, and Na\(^+\) interacts with the channels with a kinetics described by the noncooperative binding to two, kinetically distinguishable, sites.

A preliminary account of this work has been presented in abstract form (Gómez-Lagunas, 2000).

**MATERIALS AND METHODS**

**Cell Culture and Channel Expression**

Insect Sf9 cells kept in culture in Grace’s media (GIBCO BRL) at 27°C were infected, with a multiplicity of infection of 10, with a recombinant baculovirus, Autographa californica nuclear polyhedrosis virus, containing the cDNA of Shaker B, as previously reported (Klaiber et al., 1990; Gómez-Lagunas, 1997). Experiments were performed 48 h after infection.

**Electrophysiology**

Macroscopic currents were recorded under whole-cell patch clamp, with an Axopatch-1D (Axon Instruments). The currents were filtered at 5 kHz with the built in filter of the amplifier, and sampled at 100 µs/point, with a TL1 interface (Axon Instruments). Electrodes were pulled from borosilicate glass (KIMAX 51) to a final resistance of 1-2 MOhm ~80% of the series resistance was electronically compensated. Activating pulses (unless otherwise indicated = +20 mV/30 ms) in 0-K\(^+\) solutions, were delivered at 1 Hz. This procedure will be referred to as pulsing.

**RESULTS**

**Na\(^+\) Ions Speed the Collapse of G\(_K\) in 0 K\(^+\) Conditions**

With Na\(^+\)-containing, 0-K\(^+\) solutions on both sides of the membrane (Na\(_o\)/Na\(_i\); MATERIALS AND METHODS), the delivery of depolarizing pulses that activate the channels followed by repolarization to a negative potential (usually the HP = −80 mV) collapse the Shaker B K\(^+\) conductance (G\(_K\); INTRODUCTION). Fig. 1 shows that Na\(^+\) ions foster the collapse of G\(_K\). Fig. 1 A illustrates the collapse-recovery cycle of G\(_K\) in Na\(_o\)/Na\(_i\) solutions. The traces show inward K\(^+\) currents (I\(_K\)), evoked by +20 mV/30-ms activating pulses, in K\(_o\)/Na\(_i\). The left trace (Fig. 1 A, before) is a control I\(_K\), recorded at the beginning of the experiment. The middle trace (Fig. 1 A, After) is the current left after the delivery of 20 activating pulses (a procedure hereafter referred to as pulsing) while the cell was bathed in the test Na\(_o\)/Na\(_i\) solutions (not shown). After pulsing in 0 K\(^+\), Na\(^+\)-containing solutions, the channels become reluctant to conduct K\(^+\). The right trace (Fig. 1 A, Recovery) shows the recovery of I\(_K\) brought about by a 3-min depolarization to 0 mV (Gómez-Lagunas, 1997).

In contrast to the dramatic effect of pulsing in Na\(_o\)/Na\(_i\), the delivery of the same number of pulses in NMG-
containing, 0 K\textsuperscript{+}, and 0 Na\textsuperscript{+} (not added) solutions scarcely affects G\textsubscript{K}. This is shown in Fig. 1 B, which presents two K\textsuperscript{+} currents recorded in K\textsubscript{o}/NMG\textsubscript{i}, before (left trace) and after (right trace) pulsing in NMG\textsubscript{o}/NMG\textsubscript{i} (not shown). There was only a 13% reduction of I\textsubscript{K}.

Fig. 1 C compares the reduction of G\textsubscript{K} after pulsing in a variety of solutions. Notice that, whereas in Na\textsubscript{o}/Na\textsubscript{i} (last bar) G\textsubscript{K} is basically erased (99 ± 1% reduction, n = 4), in NMG\textsubscript{o}/NMG\textsubscript{i} (second bar) G\textsubscript{K} drops only 20 ± 2% (n = 16). Thus, pulsing in 0 Na\textsuperscript{+}, NMG-containing, solutions collapses G\textsubscript{K}, but far less than in the presence of Na\textsuperscript{+} ions (in fact, in Na\textsubscript{o}/Na\textsubscript{i}, only 10–15 pulses are needed to completely eliminate G\textsubscript{K}, see Gómez-Lagunas, 1997, 1999). Moreover, after pulsing in NMG most of the channels (=80%) remain in a state from which they readily collapse upon the addition of Na\textsuperscript{+} (not shown). The third bar in Fig. 1 C, shows that external choline, another impermeant and nonblocking cation, is as inert as NMG (G\textsubscript{K} drop = 21 ± 6%, n = 5; see DISCUSSION).

In summary, the combined condition, presence of Na\textsuperscript{+} and lack of K\textsuperscript{+}, makes G\textsubscript{K} more liable to collapse. Finally, notice that pulsing with Na\textsuperscript{+} ions present (added) in only the external solution (in Na\textsubscript{o}/NMG\textsubscript{i}, first bar) drops G\textsubscript{K} in about the same extent (90 ± 4%, n = 9) as it does with Na\textsuperscript{+} on both sides of the membrane (99 ± 1%, last bar). So, even when under physiological conditions, external Na\textsuperscript{+} neither permeates nor blocks Kv channels (but see Block and Jones, 1996), it effectively fosters the collapse of G\textsubscript{K} in 0-K\textsuperscript{+} solutions.
External Na⁺ Interaction with Shaker Channels

The interaction of external Na⁺ with Shaker was further studied by looking at the extent of Gₖ collapse produced by pulsing in solutions of variable [Naᵦ⁺], with NMGᵢ as the internal solution (in (NMGᵢ + [Naᵦ⁺])/NMGᵢ, see MATERIALS AND METHODS). Fig. 2 A shows that as [Naᵦ⁺] increases, Gₖ drops following a Hill saturation curve (line through the points, labeled 0 K⁺), with a Kᵦ = 3.3 mM, and maximal extent of collapse = 1.05 (Hill number n = 0.97). When the same measurements are done in the presence of either 0.08 or 0.3 mM Kᵦ, it is seen that the apparent Kᵦ for Naᵦ⁺ increases to either 4.7 or 5.3 mM, respectively, without a significant change in the maximal extent of collapse (0.98 or 0.91 with 0.08 or 0.3 mM Kᵦ, respectively). This indicates that K⁺ protection (reduction of Gₖ drop) is more efficient at the lower [Naᵦ⁺], and thus shows that K⁺ inhibits in a competitive manner the binding of Na⁺, to the site where Gₖ is modulated. The inset shows the plot in an expanded [Naᵦ⁺] scale (B). Double reciprocal plot of the points in A. 0 K⁺: r = 0.9996; 0.08 K⁺: r = 0.9868; 0.3 K⁺: r = 0.997 (see text for details). (C). Percentage of Gₖ collapse as a function of the HP during pulsing in (NMGᵢ + [Naᵦ⁺])/NMGᵢ with [Naᵦ⁺] = 2.5 mM. (D). Gₖ drop as a function of the pulse potential (Vp), during pulsing from the HP of ~80 mV, [Naᵦ⁺] = 2.5 mM as in C. The points are the mean ± SEM of at least four measurements at each [Naᵦ⁺].

Figure 2. Gₖ drop as a function of the external [Naᵦ⁺].
(A). Extent of Gₖ reduction [1/(1/Vp)], like in Fig. 1 C, after pulsing in either (NMGᵢ + [Naᵦ⁺])/NMGᵢ, 0-K⁺ solutions, with the indicated [Naᵦ⁺] (●), or in the same solutions but with 0.08 (■) or 0.3 mM Kᵦ⁺ (▼). The lines are the fit of the points with a Hill equation with parameters (●): n = 0.97, Kₒ = 3.3 mM, and maximal collapse (m.c) = 1.05; and (■): n = 0.91, Kₒ = 4.7 mM, m.c = 1.02; (▼): n = 1.10, Kₒ = 5.3 mM, m.c = 0.97. The inset shows the plot in an expanded [Naᵦ⁺] scale (B). Double reciprocal plot of the points in A. 0 K⁺: r = 0.9996; 0.08 K⁺: r = 0.9868; 0.3 K⁺: r = 0.997 (see text for details). (C). Percentage of Gₖ collapse as a function of the HP during pulsing in (NMGᵢ + [Naᵦ⁺])/NMGᵢ with [Naᵦ⁺] = 2.5 mM. (D). Gₖ drop as a function of the pulse potential (Vp), during pulsing from the HP of ~80 mV, [Naᵦ⁺] = 2.5 mM as in C. The points are the mean ± SEM of at least four measurements at each [Naᵦ⁺].
However, it is pertinent to mention that considering the possibility of ion permeation through the channels, deviations from the equilibrium conditions, on which the saturation Michaelis-Menten or Hill equations are based (Segel, 1993), might be expected, particularly at the higher $[K^+/H^+]$, so both the $K^+/H^+$ and $Na^+/H^+$ $K_d$s are likely to be overestimations of their true equilibrium values.

If the site where $Na_o^+$ (and $K_o^+$) binds was located within the electric field of the membrane, it would be expected that pulsing from a hyperpolarized HP should increase the drop of $G_K$, by favoring the $Na_o^+$ occupancy of the site.

In contrast to the above prediction, Fig. 2 C shows that, with a nonsaturating $[Na_o^+] = 2.5$ mM, the drop of $G_K$ is slightly, although significantly, reduced ($P < 0.05$), instead of increased, by pulsing from a more negative HP (from $49 \pm 4\%$ [$n = 7$] at HP $-80$ mV; to $37 \pm 1\%$ [$n = 6$] at HP $-120$ mV). On the other hand, Fig. 2 D shows that within the range of 0 to $+60$ mV, the pulse potential (Vp) does not play a significant role on the extent of collapse. The dependence on holding potential of $Na_o^+$ action is qualitatively equivalent to that of $Ba^{2+}$ protection (Gómez-Lagunas, 1999). The latter decreases by pulsing from hyperpolarized HPs, and, like $Na_o^+$ action, it is not dependent on $Vp$. The following nonexcluding possibilities had been proposed as an explanation of this voltage dependence (Gómez-Lagunas, 1999): (1) hyperpolarized HPs might promote ion ($Na_o^+$ or $Ba^{2+}$) permeation through the channels, thus reducing the strength of their corresponding effects; or (2) there could be more than one state where the state of $G_K$ is modulated, a negative HP might favor the occupancy of innermost and least effective site, in terms of the corresponding effect of the ions (see Fig. 6). Whatever the case, the effect of voltage on $Na_o^+$ and $Ba^{2+}$ actions, along with the previous finding that the potency with which ions other than $K^+$ protect $G_K$, is not related to the permeability sequence of the channels (Gómez-Lagunas, 1997, 1999) suggests that the external site, whose state of occupancy deter-
mines the state of $G_K$ (available or reluctant), may not be part of the selectivity filter, but instead that it might be externally located to it (see Discussion).

**Internal Na$^+$ Interaction with Shaker Channels**

It is known that internal Na$^+$ interacts with the pore of K channels blocking $I_K$ (Bezanilla and Armstrong, 1972; French and Wells, 1977; Marty, 1983; Yellen, 1984; Neyton and Pellechi, 1991; Starkus et al., 2000). To see if the collapse of $G_K$ occurs as a discontinuous change only observed in the limit of 0 K$^+$ (molar fraction of Na$^+$, $X_{Na}$ = 1), or if there is a gradual change in the properties of the channels as $X_{Na}$ increases, the behavior of the channels as the molar fraction of Na$^+$ ($X_{Na,i}$) approached 1 was studied, in internal solutions where $X_{Na,i} + X_{K,i} = 1$.

For a reference, Fig. 3 A shows a typical I-V relationship obtained under standard conditions ($Na_o/K_i$). Above 0 mV, where the probability of opening is 1, $I_K$ increases linearly with the voltage. With the $X_{Na,i} = 0$, this is always observed. An I-V obtained with a low $X_{Na,i} = 0.21$ is shown in Fig. 3 B. First $I_K$ increases with the voltage, but at about +40 mV, the current starts to deviate from the linearity and after that a region of negative conductance develops, as Na$^+$ blocks the channels. The departure from linearity, at positive voltages (+40 mV), is more clearly seen by comparing the experimental points with the straight line in the plot. The latter is the least-square fit of the points between 0 and +30 mV, and therefore gives an estimate of the expected $I_K$ if $X_{Na,i}$ was zero. Thus, the difference between the line and the points in the graph is an estimate of the extent of block at each voltage, the latter is plotted in Fig. 3 C, which presents the mean ± SEM of four cells. The line is the fit of the points with a Woodhull equation (Fig. 3 legend), with electrical distance $\delta = 0.7$, and $K_d(0 \text{ mV}) = 538.3 \text{ mM}$ (Woodhull, 1973). A representative I-V ($n = 6$ cells) obtained with a higher $X_{Na,i} = 0.42$ is shown in Fig. 3 D. See that, qualitatively, it looks like that with $X_{Na,i} = 0.21$. With the Na$^+$ solution, and $X_{Na,i}$ up to 0.42, Na$^+$ block becomes continuously stronger as the voltage is made more positive, for voltages up to +160 mV.
In contrast to the behavior with \( X_{Na,i} \approx 0.42 \), Fig. 4 A shows that with \( X_{Na,i} = 0.71 \), the IV relationship acquires an N shape (French and Wells, 1977, Starkus et al., 2000) within the voltage range \( V_p \leq +160 \text{ mV} \). \( I_k \) first increases, but starts to deviate from the linearity as \( Na^+ \) blocks the channels, and thereafter a region of negative conductance develops, until a “critical” voltage \( (V_c) \) is reached (in this case \( V_c = +70 \text{ mV} \)) where \( Na^+ \) block is relieved and the current increases again with the voltage (pointed out by the arrow), giving the IV its N shape. The traces in the right panel of Fig. 4 A illustrate the N shape of the IV relationship. See that the peak currents are in the following order: \( I (V_c = +110 \text{ mV}) > I (+30 \text{ mV}) > I (+70 \text{ mV}) \). Finally, Fig. 4 B shows an IV obtained with \( X_{Na,i} = 0.83 \). Notice that although its overall shape is like that observed with \( X_{Na,i} = 0.71 \), \( V_c \) is smaller \( (V_c = +50 \text{ mV}) \). The traces in the right panel illustrate the N shape of the IV relationship.

In the N-shaped IVs, there is a voltage \( (V_c) \) where \( Na^+ \) block is overcome (French and Wells, 1977). From the value of \( V_c \), the minimal energy required to unblock the channels \( (B_{Na}) \) can be estimated as follows (Latorre and Miller, 1983): \( B_{Na} = \exp[(B_{Na} - B_K)/RT] \). For example, with \( X_{Na,i} = 0.42 \) the IV is not N-shaped, so \( V_c \approx +160 \text{ mV} \), then when \( X_{Na,i} \) increases to for example 0.83, \( B_{Na} \) changes from at least 15.5 \( (X_{Na,i} = 0.42 \), underestimating \( V_c \) as \( +160 \text{ mV} \)) to 5.5 \( \pm 0.5 \text{ kJ/mol} \) \( (X_{Na,i} = 0.83) \) that is 10 \text{ kJ/mol} \), whereas \( \Delta \mu_{Na} \) changes from at least 12.9 to 4.0 \text{ kJ/mol} \), that is 8.9 \text{ kJ/mol} \.

By looking at \( Na^+ \) block of the squid K channel, N-shaped IVs were first observed by French and Wells (French and Wells, 1977), whom after observing that block was overcome by very positive potentials, proposed that the relative permeability of \( Na^+ \) to that of \( K^+ \) \( (P_{Na}/P_K) \) increased at those voltages (more than \( +160 \text{ mV} \)). This proposal has received support from a recent study of the change of the \( P_{Na}/P_K \) ratio of Shaker channels, during the onset of slow inactivation (Starkus et al., 2000). In the simple case where a single, rate limiting, barrier B limits ion permeation through a channel, the relative permeability of a test ion \( (Na^+) \) to that of a reference ion \( (K^+) \) can be estimated with the relation (Reuter and Stevens, 1980; Latorre and Miller, 1983): \( P_{Na}/P_K = \exp[(B_{Na} - B_K)/RT] \), where \( R \), \( T \) have their usual meanings.

Following this simplifying approach, and taking \( B_K = 0 \), and \( B_{Na} \) as in Fig. 5 A, it is seen that when \( X_{Na,i} \) increases from 0.71 to 0.83 (1.2 times), \( P_{Na}/P_K \) increases about four times (Fig. 5 B). The results in Figs. 3–5 show that as \( X_{Na,i} \) increases there is an actual reduction (i.e., a reduction not entirely accounted for by the change in \( \Delta \mu_{Na} \)) of the energy required to relieve \( Na^+ \) block, yielding a substantial change of the \( P_{Na}/P_K \) ratio.

The above effect could be explained as the result of an increase in the electrostatic repulsion between the blocking \( Na^+ \) and a neighbor ion, as \( X_{Na,i} \) increases. However, it is not easy to see why this repulsion should be bigger as the molar fraction of the permeant \( K^+ \) ion decreases. Alternatively, it could be that as \( X_{Na,i} \) increases, there is a deformation of the pore, measured as a reduction of the energy required to allow \( Na^+ \) permeation. If that were
the case, it could be that this deformation were maximal in the limit where \( X_{\text{Na},i} = 1 \). Pulsing under the latter conditions (\( \text{Na}_o/\text{Na}_i \)) collapses \( G_K \) (Gómez-Lagunas, 1997). Finally, as aforementioned (Fig. 2), the voltage dependence of external \( \text{Na}^+ \) and \( \text{Ba}^{2+} \) actions, as well as that of internal \( \text{Ba}^{2+} \) (Gómez-Lagunas, 1999), might be explained by the involvement of two sites in the drop of \( G_K \); thus, to explore this point, the role of internal \( \text{Na}^+ \) on the fall of \( G_K \), when \( X_{\text{K},i} = 0 \), was studied.

Fig. 6 A shows that pulsing with \( \text{Na}^+ \) present (added) in only the internal solution (\( \text{NMG}_o/\text{Na}_i \)), drops \( G_K \) in about the same extent (93 ± 5%, \( n = 6 \)) as it does with \( \text{Na}^+ \) present on both sides of the membrane (last bar, 99 ± 1%, \( n = 4 \)). For comparison, the figure also shows the collapse in \( \text{NMG}_o/\text{NMG}_i \), as in Fig. 1. The above observation suggests that \( \text{Na}^+ \) could possibly act on the same site regardless of the side of the membrane from where it comes. To further explore this point the collapse of \( G_K \) as a function of \( [\text{Na}_i^+] \) (in \( \text{NMG}_o/(\text{NMG}_i-[\text{Na}_i]) \)) was determined. Fig. 6 B shows that as \( [\text{Na}_i^+] \) increases \( G_K \) drops following a Hill curve (line through the points) with \( n = 1.16 \) (which indicates that there is, at least, one site where \( \text{Na}^+ \) binds) plus an offset, that accounts for the drop in, \( 0-\text{Na}^+ \), \( \text{NMG} \) solutions. The points are the mean ± SEM of at least four cells at each \( [\text{Na}_i^+] \). (C) Double reciprocal plot of the points in B. The two dotted lines are the least-squares fit of the points, on them, with the following parameters: \( r = 0.994, K_{\text{d}1} = 2 \text{ mM}, \text{maximal collapse} (\text{m.c})_1 = 0.6; \) and: \( r = 0.970, K_{\text{d}2} = 14.3 \text{ mM}, (\text{m.c})_2 = 0.36 \). The inset shows the Eadie-Scatchard plot of the points in B. (D) \( G_K \) drop as a function of the pulse potential, during pulsing in \( \text{NMG}_o/(\text{NMG}_i-[\text{Na}_i]) \), with \( [\text{Na}_i^+] = 10 \text{ mM} \).

![Figure 6](https://example.com/figure6.png)

**Figure 6.** \( G_K \) drop in, 0 K+, \( \text{Na}^+ \)-containing internal solutions. (A). \( G_K \) reduction after pulsing with the cell bathed in either \( \text{NMG}_o/\text{NMG}_i \), \( \text{NMG}_o/\text{Na}_i \) or \( \text{Na}_o/\text{Na}_i \) solutions, as indicated. (B) \( G_K \) reduction, \( 1 - (I/I_o) \), as a function of \( [\text{Na}_i^+] \). \( I \) is the control \( I \) at +20 mV, and \( I_o \) is the current left after pulsing with the cell in \( \text{NMG}_o/(\text{NMG}_i-[\text{Na}_i]) \) with the indicated \( [\text{Na}_i^+] \), like in Fig. 2 (Materials and Methods). The line is the fit of the points with a Hill equation with \( n = 1.16 \) and an offset = 0.2 that accounts for the drop in, 0-\( \text{Na}^+ \), \( \text{NMG} \) solutions. The points are the mean ± SEM of at least four cells at each \( [\text{Na}_i^+] \).
and Ba\textsuperscript{2+} effects, the voltage dependence of Na\textsuperscript{+} and Ba\textsuperscript{2+} actions are different. The latter decreases markedly with hyperpolarized HPs, and it is not dependent on Vp (Gómez-Lagunas, 1999). Further work is needed to understand this difference.

**DISCUSSION**

The Shaker B G\textsubscript{K} collapses when the channels close in 0-K\textsuperscript{+} solutions (Gómez-Lagunas, 1997, 1999; Melishchuk et al., 1998). Prolonged depolarizations are needed to recover G\textsubscript{K} (Gómez-Lagunas, 1997). The drop of G\textsubscript{K} seems to involve major changes in the conformation of the channels including not only the pore (Gómez-Lagunas, 1997), but also the voltage sensor (Melishchuk et al., 1998). So it could be that recovery is so slow due to the stability of the collapsed conformation, and to the number of rearrangements needed to reset the channels in their normal conducting state.

Here, it was shown that the combined condition 0 K\textsuperscript{+} and presence of Na\textsuperscript{+} makes G\textsubscript{K} more liable to collapse, as Na\textsuperscript{+} ions speed (in number of pulses) the drop of G\textsubscript{K}.

Nonetheless, Na\textsuperscript{+} is not necessary for G\textsubscript{K} to collapse, as it also falls in NMG or choline solutions. Considering that Na\textsuperscript{+} binds with millimolar affinity from both sides of the membrane, it seems unlikely that the drop of G\textsubscript{K} in the 0-Na\textsuperscript{+} solutions, could be produced by contaminant Na\textsuperscript{+} ions. More likely, the binding of Na\textsuperscript{+}, instead of K\textsuperscript{+}, or just the absence of K\textsuperscript{+} in the pertinent site(s) when the channels close, elicits the drop of G\textsubscript{K}, although with significantly different speeds that might be accounted for by a smaller dwelling time of K\textsuperscript{+}, in the pertinent site(s), in the presence of Na\textsuperscript{+} ions than in their absence, in agreement with the observed mutually exclusive binding of K\textsuperscript{+} and Na\textsuperscript{+} ions to the externally located site.

**Extracellular Na\textsuperscript{+} and the Drop of G\textsubscript{K}**

Under physiological conditions, Na\textsubscript{o}\textsuperscript{+} neither permeates nor blocks Kv channels (but see Block and Jones, 1996), however, it effectively fosters the drop of G\textsubscript{K}, by binding to a site with an apparent K\textsubscript{d} = 3.3 mM. External Na\textsuperscript{+} and K\textsuperscript{+} compete, in a mutually exclusive manner, for binding to this site, and from this observation a high, micromolar affinity for K\textsuperscript{+} was estimated, K\textsubscript{d} = 80 μM. The presence of high affinity binding sites for K\textsuperscript{+} in the selectivity filter of the channels, from where, under physiological conditions, Na\textsuperscript{+} is excluded because of its lower affinity, has been shown to underlie the structural basis of the mechanism of selectivity of K channels (e.g., Korn and Ikeda, 1995; Doyle et al., 1998; Ogiełksa and Aldrich, 1998).

Could the site where K\textsuperscript{+} and Na\textsuperscript{+} bind modulating G\textsubscript{K} be located within the selectivity filter? The protection exerted by TEA\textsubscript{o} suggests that the site is externally located (Gómez-Lagunas, 1997), as the selectivity filter is (Doyle et al., 1998). Nonetheless, the voltage dependence of Na\textsuperscript{+}, Ba\textsuperscript{2+}, and K\textsuperscript{+} actions (Gómez-Lagunas, 1999; Fig. 2), along with the observation that the potency with which monovalent cations protect is not related to the selectivity of the channels (Gómez-Lagunas, 1997), together suggest that this site could be external to the selectivity filter.

However, considering that in multi-ion pores, the voltage dependence of ligand binding depends on ion occupancy (Neyton and Miller, 1988a,b; Spassova and Lu, 1999; Thompson and Begenisch, 2001), and that selectivity of Kv channels depends on the [K\textsuperscript{+}] (Korn and Ikeda, 1995; Ogiełksa and Aldrich, 1998; Fig. 5), the selectivity filter cannot yet be ruled out as the externally located place where G\textsubscript{K} is modulated (Melishchuk et al., 1998). Although, the drop of G\textsubscript{K} with [Na\textsubscript{o}\textsuperscript{+}] follows a Hill equation with n = 0.97 and a linear double reciprocal plot, the reduced effectiveness of Na\textsuperscript{+} at hyperpolarized HPs, suggested the involvement of more than one site in the collapse of G\textsubscript{K}. This possibility was reinforced by the kinetics of the drop of G\textsubscript{K} with [Na\textsubscript{o}\textsuperscript{+}] (see below).

**Internal Na\textsuperscript{+} Interaction with Shaker**

For voltages up to +160 mV, I-V relationships obtained with the Na\textsubscript{o} solution, and with a variable X\textsubscript{Na\textsubscript{i}}, have a shape that depends on X\textsubscript{Na\textsubscript{i}}. With X\textsubscript{Na\textsubscript{i}} ≥ 0.71, the I-Vs are N-shaped. In contrast to the behavior of Shaker channels, French and Wells (1977) did not find a significant change of Vc with the internal Na\textsuperscript{+} in the squid K channel. It would be interesting to determine if the differential dependence of the I-V relationships on X\textsubscript{Na\textsubscript{i}} of these channels, keeps any relation to their different behavior in 0-K\textsuperscript{+} solutions (Almers and Armstrong, 1980; Khodakhah et al., 1997; Gómez-Lagunas, 1997, 1999).

Recently, by looking at the onset of the slow inactivation of Shaker channels lacking the NH\textsubscript{2} terminus domain, Starkus et al. (2000) afforded evidence showing that once Vc is reached and the conductance becomes again positive, Na\textsuperscript{+} actually flows throughout the channels, carrying current from the internal to the external solution. This supports the proposal of French and Wells (1977) that selectivity is voltage-dependent. The results presented here show that the Shaker selectivity also varies with X\textsubscript{Na\textsubscript{i}}. It is pertinent to point out that an analogous statement has been made for the squid Na\textsuperscript{+} channel (Cahalan and Begenisch, 1976).

It has been shown that mammalian Kv2.1 channels as well as DR channels of bullfrog neurons although remain stable in 0 K\textsuperscript{+}, conducting Na\textsuperscript{+}, undergo a conformational change of the pore region, which is observed as a reduction of the capability of TEA to block the channels (Korn and Ikeda, 1995; Block and Jones, 1997). Likewise, it seems that as X\textsubscript{Na\textsubscript{i}} increases there is a deformation of the Shaker pore, which is observed as a reduction of the energy barrier that limits Na\textsuperscript{+} efflux.
That deformation could be maximal when $X_K = 0$, but further studies are needed to determine its relation with the collapse of $G_K$ here studied.

When $X_{K,i} = 0$, pulsing with Na$^+$ ions added to the internal solution alone drops $G_K$ in about the same extent as it does with Na$^+$ on both sides of the membrane, suggesting that Na$^+$ might be able to reach the externally located site where $G_K$ is modulated, in agreement with the observation that Vc decreases as $X_{K,i}$ approaches one, and with reports showing Na$^+$ currents through $K_v$ channels in 0 K$^+$ (Zhu and Ikeda, 1993; Korn and Ikeda, 1995; Block and Jones, 1997; Starkus et al., 1997, 2000; Melishchuk et al., 1998; Ogieska and Aldrich, 1998; Zhuren et al., 2000). On the other hand, the [Na$^+$] dependence of $G_K$ drop indicates that there are actually two binding sites, of which the one with the higher affinity ($K_a = 2$ mM) could be the externally located site ($K_a = 3$ mM). It might be that presence of two sites could form the structural basis of the $X_{Na,i}$ dependence of Vc. Whatever the case, it is pertinent to mention that the possible involvement of two sites in the collapse of $G_K$ was previously suggested by the characteristics of the protection afforded by internal Ba$^{2+}$ (Gómez-Lagunas, 1999).

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